The role of adipose tissue immune cells in obesity and low-grade inflammation

Milos Mraz and Martin Haluzik

Third Department of Medicine – Department of Endocrinology and Metabolism, General University Hospital, First Faculty of Medicine of Charles University in Prague, U nemocnice 1, 128 00 Prague 2, Czech Republic

Correspondence should be addressed to M Mraz Email milos_mraz@yahoo.co.uk

Abstract

Adipose tissue (AT) lies at the crossroad of nutrition, metabolism, and immunity; AT inflammation was proposed as a central mechanism connecting obesity with its metabolic and vascular complications. Resident immune cells constitute the second largest AT cellular component after adipocytes and as such play important roles in the maintenance of AT homeostasis. Obesity-induced changes in their number and activity result in the activation of local and later systemic inflammatory response, marking the transition from simple adiposity to diseases such as type 2 diabetes mellitus, arterial hypertension, and ischemic heart disease. This review has focused on the various subsets of immune cells in AT and their role in the development of AT inflammation and obesity-induced insulin resistance.

Key Words

- ▶ adipose tissue
- immune cells
 - low-grade inflammation
- ▶ innate immunity
- adaptive immunity
- ▶ insulin resistance

Journal of Endocrinology (2014) 222, R113–R127

Introduction

Obesity is nowadays considered the pandemia of the 21st century and its rapidly growing prevalence together with associated complications comprises one of the gravest healthcare problems of our society (O'Rahilly 1997, York *et al.* 2004). According to WHO, in 2011 ~500 million people worldwide suffered from obesity (BMI being $> 30 \text{ kg/m}^2$) and these numbers are estimated to at least double until 2030 (http://www.who.int/mediacentre/fact-sheets/fs311/en/). The main burden of obesity lies in its interconnection with a number of metabolic and non-metabolic diseases including type 2 diabetes mellitus (T2DM), dyslipidemia, arterial hypertension, and atherosclerosis, leading to substantially increased cardiovascular and cerebrovascular morbidity and mortality.

As excessive accumulation of body fat (mostly due to the imbalance between energy intake and expenditure) lies at the core of all these problems, it is specifically the adipose tissue (AT) that plays the pivotal role in the

http://joe.endocrinology-journals.org DOI: 10.1530/JOE-14-0283 development of obesity-related complications. AT is not any more considered a mere storage site for excessive energy or a means of thermal and mechanical isolation as it was some 20 years ago. Instead, years of intense research have brought upfront a picture of a highly active organ involved in numerous metabolic, hormonal, and immune processes, whose products and reactions are able to act not only locally but influence also other organs and systems and play a crucial role in the whole-body homeostasis. Several mechanisms as to how increased amounts of AT may lead to metabolic derangements and enhanced atherosclerosis have been proposed, including endocrine dysfunction (Bluher 2009), AT hypoxia (Trayhurn 2013), or decreased lipid storage capacity with their subsequent ectopic accumulation (Ravussin & Smith 2002). However, one of the most promising concepts integrating excess adiposity with T2DM and cardiovascular complications includes the development of local and systemic chronic

low-grade inflammation characterized by increased infiltration of immune cells into AT and increased production and subsequent secretion of proinflammatory factors into circulation (Neels & Olefsky 2006). This review has focused on the various subsets of immune cells in AT and their role in the development of low-grade inflammation and insulin resistance (IR).

Metabolism, immunity, and inflammation

Inflammation is a series of cellular and humoral reactions aimed at defending the body from various insults including infection and tissue damage and leading ultimately to the restoration of functional and morphological integrity of affected tissues (Cildir et al. 2013, Lee & Lee 2014). Typically, in acute inflammation, the initial damaging insult triggers the release of a number of immunomodulatory molecules including cytokines and chemokines from tissue-resident macrophages and mast cells, provoking a rapid recruitment of neutrophils first and then macrophages and lymphocytes from circulation to the inflammation site (Cildir et al. 2013). The infiltrating cells then destroy the infectious agents and remove damaged cells. Finally, the transition from innate to adaptive immunity is performed via antigen-presenting cells (APC) and B- and T lymphocytes. In general, inflammation is characterized by increased local and systemic cytokine levels along with increased number of infiltrating immune cells, with neutrophils dominating mainly in acute phases while macrophages take the stage in more chronic conditions (Lee & Lee 2014).

Obesity was shown to be associated with a slightly different type of inflammation referred to as chronic lowgrade sterile inflammation or metainflammation (inflammation in metabolic tissues) and characterized by only a modest increase in circulating proinflammatory factors and the absence of clinical signs of inflammation (hence the term subclinical inflammation) (Medzhitov 2008). Despite its much lower intensity (as compared with e.g. sepsis as a model of hyperacute generalized inflammatory response), obesity-induced inflammation exerts profound effects on metabolic pathways, playing one of the central roles in the development of IR (Heilbronn & Campbell 2008, Oliver et al. 2010). Although a connection between inflammation and T2DM was suggested more than a century ago with first attempts to treat hyperglycemia by anti-inflammatory drugs (Williamson 1901), the evidence for causal relationship between inflammation and IR started to emerge some 25 years ago, when it was shown by Feingold et al. (1989) that the

administration of the chief proinflammatory cytokine tumor necrosis factor α (TNF α) resulted in increased serum glucose concentrations. Subsequently, Hotamisligil et al. (1993) found that TNFa was elevated in obese rodents and that its neutralization by specific antibodies markedly improved insulin sensitivity. Moreover, TNFa knockout leads to improved insulin sensitivity in diet-induced obesity (Uysal et al. 1997). The link between metabolism and immunity was further corroborated at the intracellular level, with findings that the main inflammatory signaling pathway comprising nuclear factor-kB (NF-kB) and inhibitor of κ B kinase- β (IKKB) is stimulated in obesity as well as in IR (Shoelson et al. 2003). Conversely, genetic deletion of IKKB or the inhibition of this pathway by salicylates attenuated IR in both mice and humans (Yuan et al. 2001, Shoelson et al. 2003). Furthermore, obesityrelated inflammation tends to activate also other proinflammatory factors including the group of c-Jun N-terminal protein kinases (JNK), while the ablation of JNK protects experimental animals from diet-induced obesity and inflammation (Hirosumi et al. 2002, Solinas et al. 2007). Collectively, these findings on the tight interconnection of metabolism, immunity, and inflammation have given rise to an entirely new field of biomedical research referred to as immunometabolism.

AT as immune organ

The morphological and functional proximity of immune and metabolic reactions can be traced back in evolution to first multicellular animal species. In insects, an organelle termed fat body, which contains a receptor for bacterial and fungal antigens (Toll receptor), is responsible for innate immunity (Leclerc & Reichhart 2004). Toll receptor activates the signaling cascade of NFKB leading to the secretion of antimicrobial peptides and activation of further defense mechanisms (Rolff & Siva-Jothy 2003). At the same time, the fat body functions as a metabolic organ and storage site for lipids (Sondergaard 1993, Rusten et al. 2004). In vertebrates, these functions were divided between liver, AT, and bone marrow. Even though formerly only bone marrow and liver (especially after the discovery of acute-phase proteins) were associated with immunity reactions, recent data have unequivocally shown that AT also maintains at least a part of the functions of an immune organ (Mortensen 2001).

From the histological point of view, AT is composed of two distinct entities – adipocytes (mature fat cells) and the interadipocytar stromal-vascular fraction formed by extracellular matrix with dispersed fibroblasts, preadipocytes

http://joe.endocrinology-journals.org DOI: 10.1530/JOE-14-0283

(immature adipocyte precursors), endothelial, and immune cells (Curat et al. 2004). AT-resident immune cells include almost the full spectrum of immune cell types, playing important roles in tissue housekeeping, removal of detritus, and apoptotic cells, and tissue homeostatis maintenance, under non-obese conditions (Schipper et al. 2012a). However, excessive fat accumulation leads to substantial changes in the amount and function of immune cells increasing the number and activity of some of them (most notably macrophages, mast cells, neutrophils, and T- and B lymphocytes) while simultaneously reducing others including eosinophils and several subsets of T lymphocytes (T helper 2 (Th2), Treg, and iNKT cells) (Cildir et al. 2013). This imbalance lies at the very core of the development of obesity-related local and systemic inflammation (Table 1).

Interestingly, AT inflammation accompanies not only accumulation of body fat but also its rapid reduction induced, e.g. by short-term caloric restriction or in the first weeks after a bariatric procedure (Mraz *et al.* 2011, Trachta *et al.* 2014). Moreover, subjects with severely depleted fat reserves as seen in mental anorexia also show increased production of proinflammatory adipokines (Dolezalova *et al.* 2007). These data suggest that rapid or extreme changes in body fat content provoke immune response

regardless of their direction. Other factors than changes in body weight can also contribute to the development of AT inflammation, including acute (e.g. major surgery) as well as chronic conditions (e.g. end-stage renal disease) (Kremen et al. 2006, Roubicek et al. 2009). More recently, gut microbiota has been identified as an important modifier of local as well as systemic inflammatory reactions influencing, except of the intestine, also remote tissues, most notably peripheral blood and AT, especially its visceral compartment, where, via the portal vein, intestinal microbial products are being directly drained into (Burcelin et al. 2013). Obesity and T2DM were associated with changes in the amount and composition of gut microbes in experimental animals as well as humans (Carvalho & Saad 2013, Cox & Blaser 2013). Interestingly, different gut microbiota-derived products can exert both pro- and antiinflammatory effects, as e.g. the translocation of several gut microbial antigens (mainly lipopolysaccharide and peptidoglycans) into systemic circulation leads to metabolic endotoxemia, suggested as one of the main triggers of AT and systemic low-grade inflammation (Burcelin et al. 2013, Carvalho & Saad 2013). In contrast, the products of gut bacterial fermentation of ingested dietary fiber, especially short-chain fatty acids (SCFA) - mainly butyrate, propionate, and acetate - were shown to have anti-inflammatory

Immune cell type	Antigens and other markers	Main secretory products		
			Relationship with insulin resistance	
Myeloid cells				
Macrophages				
M1	F4/80, CD11b, CD11c	INFα, IL6, NOS2	Ť	
M2	CD206, CD209, CD301, LYVE1	IL10, IL1Ra, arginase 1	\downarrow	
Dendritic cells	CD1c, CD11c, CD80, CD83, CD86	IL12, IL15	<u>↑</u>	
Mast cells	CD117, FCER1	Histamine, PGE ₂ , LTB4, TNF α , IL1 β , IL6, TGF β , IL4, IL10	<u>↑</u>	
Neutrophils	CD66b, CD11b, Ly6g	Lysozyme, NE, MPO, TNFα, IL1β, IL8, MIP1α	1	
Eosinophils	CD45, Siglec8	IL4, IL10, IL13, TGFβ	Ţ	
Lymphoid cells				
T lymphocytes				
Helper (Th)				
Th1	CD4	IENv	↑	
Th2	CD4			
Th17	CD4		↓	
Trea	CD4 $CD25$ Eoxp3			
Cytotoxic	CD4, CD23, 10xp3	Perforines granzymes IEN ₂	↓ ↑	
Notural killer T			1	
Natural Killer I		ΠΝΓα, ΙΓΙΝΥ, ΙL4, ΙL13		
B lymphocytes			Ť	
innate lymphoid type 2 cells	CD25	IL5, IL13	↓	

 \uparrow , associated with increased insulin resistance; \downarrow , associated with decreased insulin resistance; CD, cluster of differentiation; FCER1, high-affinity IgE receptor; Foxp3, forkhead box P3; IL, interleukin; IL1Ra, interleukin 1 receptor antagonist; IFNγ, interferon gamma; IgG2c, immunoglobulin G2c; LTB4, leukotriene B4; Ly6g, lymphocyte antigen 6g; LYVE1, lymphatic vessel endothelial hyaluronan receptor 1; MPO, myeloperoxidase; NE, neutrophil elastase; NK1.1, natural killer antigen 1.1; NOS, nitric oxide synthase; PGE₂, prostaglandin E₂; Siglec, Sialic acid-binding Ig-like lectin; TGF β , transforming growth factor beta; TNF α , tumor necrosis factor alpha.

http://joe.endocrinology-journals.org DOI: 10.1530/JOE-14-0283 © 2014 Society for Endocrinology Printed in Great Britain

effects and influence energy homeostasis (Kim *et al.* 2014). By acting through the G-protein-coupled receptors 41 (GPCR41) and 43 (GPCR43) that are abundantly found in AT, as well as on immune cells including peripheral blood mononuclear cells, eosinophils, and neutrophils, SCFAs (especially butyrate) were able to reduce chemotaxis and cell adhesion and thus at least partially prevent infiltration of immune cells into AT (Meijer *et al.* 2010, Kim *et al.* 2014). Treatment with propionate reduced proinflammatory cytokine and chemokine secretion from human AT as well as from macrophages (Al-Lahham *et al.* 2012). Moreover, SCFAs were also shown to inhibit the activation and proliferation of T cells and adhesion of APCs contributing further to their inflammation-reducing properties (Meijer *et al.* 2010).

Immune cells are generally categorized into two lines according to their maturation site - the myeloid line includes macrophages, dendritic cells (DCs), mast cells, and granulocytes (neutrophils, eosinophils, and basophils), while the lymphoid line consists of T- and B lymphocytes, natural killer (NK) cells, and natural killer T (NKT) cells (Kondo et al. 2003). Myeloid cells are considered the main players in innate immunity and as macrophages are the most abundant immune cell type in AT and their infiltration forms the basis of AT inflammation, innate immunity was long considered the sole immunity type involved in obesityrelated inflammation. However, several myeloid cells play important roles in the development of adaptive immunity e.g. DCs serve as antigen presenters for adaptive immunity effector cells and a number of cytokines produced by macrophages, mast cells, and neutrophils are indispensable for the activation of T- and B lymphocytes (Lee & Lee 2014). As lymphocytes are the second-largest immune cell fraction in obese AT with changes in amount and activity occurring even before the ones in macrophages, it seems that adaptive immunity also takes its turn in the processes of metainflammation (Table 1).

AT myeloid cells

AT macrophages

Macrophages are tissue-resident phagocytes that, except of serving as sentinels of innate immunity reactions, fulfill a number of housekeeping tasks (Galli *et al.* 2011). AT macrophages (ATMs) represent the largest subpopulation of AT immune cells, encompassing 5% of all cells in lean rodent AT (10–15% in the visceral depot) and rising to as much as 50% in obese animals (Weisberg *et al.* 2003, Xu *et al.* 2003). The number of ATMs in humans is lower,

http://joe.endocrinology-journals.org DOI: 10.1530/JOE-14-0283 but still comprises 4% of lean visceral fat with an increase to 12% when developing excess adiposity (Harman--Boehm *et al.* 2007). This massive infiltration of AT by ATMs together with their altered function and anatomical localization is nowadays considered the culprit of obesityrelated inflammation.

First evidence about the significance of ATM infiltration came in 2003 from the works of Xu and Weisberg who demonstrated in rodent models that obesity is associated with increased numbers of ATMs and that the majority of cytokines produced in obese AT are ATM derived (Weisberg et al. 2003, Xu et al. 2003). Subsequent studies further confirmed these findings in humans, especially in the visceral AT depot, showing that ATM content increases even more in the presence of abdominal obesity and that weight reduction is accompanied by a decrease in ATM numbers (Cancello et al. 2005, 2006, Harman-Boehm et al. 2007, Apovian et al. 2008, Vitseva et al. 2008). Moreover, ATM infiltration correlated positively not only with BMI but also with adipocyte size and stromal-vascular expression of a number of proinflammatory factors associated with IR, including TNFa, inducible nitric oxide synthase (iNOS), and IKKB (Curat et al. 2006, Lumeng et al. 2007a, Nguyen et al. 2007). In addition, a direct relationship of obese ATMs with other metabolic and non-metabolic disorders including endothelial dysfunction and non-alcoholic steatohepatitis has been established suggesting a role for ATMs in the pathogenesis of obesity-related complications that goes beyond local AT inflammation (Cancello et al. 2006, Apovian et al. 2008).

Most macrophages infiltrating obese AT come from the sources outside of body fat, mainly from systemic circulation. Studies on animals with macrophage antigen CD45.2 that were transplanted with CD45.1⁺ bone marrow showed that 85% of ATMs came from the transplanted tissue and only 15% were from the animals themselves (Weisberg et al. 2003). Nevertheless, it seems that a small fraction of ATMs can originate from local preadipocytes, as activated preadipocytes exert several antigenic characteristics similar to macrophages, including the expression of macrophage antigens F4/80, Mac1, CD80, CD86, and CD45, and are capable of phagocytosis when injected into peritoneal cavity or brought into contact with peritoneal macrophages in vitro (Charriere et al. 2003, Xu et al. 2003). These macrophage-like preadipocytes could thus comprise one of the primary cellular initiators of AT inflammation, though this hypothesis requires further confirmation.

The exact mechanisms of macrophage recruitment into the AT still remain only partially elucidated.

The candidate stimuli include a number of processes ranging from adipocyte hypertrophy and necrosis, through tissue hypoxia, lipid spillover, metabolic endotoxemia, and endoplasmatic reticulum (ER) stress to the effects of other subtypes of AT immune cells (Maury & Brichard 2010). Regardless of the initial impulse, the crucial role of attracting circulating monocytes into the tissue is played by a complex network of chemotactic cytokines (chemokines) secreted from the AT and their corresponding receptors on the attracted immune cells. Although a number of chemokines have been shown to be involved in ATM recruitment, the most promising chemotactic pathways include monocyte-chemoattracting protein 1/chemokine C-C motif receptor 2 (MCP1/CCR2), chemokine CX₃C motif ligand 1/chemokine CX₃C motif receptor 1 (CX₃CL1/CX₃CR1), and leukotriene B4/leukotriene B4 receptor (LTB4/BLT1) (Osborn & Olefsky 2012). MCP1, predominantly secreted from hypertrophic adipocytes, binds to the CCR2 receptor on macrophages stimulating thus their migration (Gerhardt et al. 2001, Christiansen et al. 2005). Overexpression of MCP1 leads to ATM infiltration, IR, and liver steatosis without increasing body weight, while deletion of MCP1 or macrophage CCR2 reduces ATM numbers in AT and improves insulin sensitivity (Kanda et al. 2006, Weisberg et al. 2006). However, other data do not fully confirm these findings (Chen et al. 2005), which can be at least partially explained by the complexity and redundancy of the chemokine network, as most chemokines are able to bind to several receptors and vice versa, and thus the blockage of one pathway might be in vivo bypassed by increased activity of a similar chemotactic pathway. Other chemokines, whose serum concentrations and AT expression are increased in obesity and to some extent correlate with insulin concentrations, include CCL3 (MIP1a-macrophage inflammatory protein 1 α), CCL5 (RANTES – regulated upon activation, normal T-expressed and secreted), CCL7 (MCP3), CCL8 (MCP2), CCL11 (eotaxin), and CCL13 (MCP4) (Hashimoto et al. 2006, Vasudevan et al. 2006, Huber et al. 2008).

Obesity alters not only the number of ATMs but also their function and tissue distribution. Based on the expression of different antigens and cytokines, macrophages can be generally divided into two subpopulation types – the classically activated M1 type and the alternatively activated M2 type. M1 macrophages can be induced *in vitro* by treating bone marrow-derived cells (BMDC) with proinflammatory cytokines including granulocyte-macrophage colony-stimulating factor (GM-CSF), IFN γ , and bacterial lipopolysaccharides resulting in a potent proinflammatory and antibacterial cell type. M2 polarization occurs under the influence of antiinflammatory IL4, IL13, and IL10 and macrophage colony-stimulating factor (M-CSF), leading to the development of a more diverse macrophage phenotype involved in anti-inflammatory and antiparasitic reactions, tissue remodeling, and wound healing (Gordon 2003, Mosser 2003).

In AT, the type of macrophage polarization depends upon the degree of adiposity. In lean individuals, ATMs, which are diffusely dispersed among adipocytes, exert predominantly a M2 phenotype expressing M2 antigens such as CD206 (mannose receptor), CD209, and CD301 (Mgl1/2); secreting anti-inflammatory IL10, IL1 receptor antagonist (IL1Ra); and the enzyme arginase 1 which blocks the activity of the proinflammatory iNOS (Chawla et al. 2011). As such, these M2-polarized ATMs fulfill a number of homeostatic functions including clearing cellular debris, regulating proliferation, and differentiation of adipocyte precursors as well as angiogenesis and thermogenesis and remodeling extracellular matrix (Chawla et al. 2011, Nguyen et al. 2011, Sun et al. 2011). The most important cytokines for M2 maintenance are IL4 originating mostly from AT eosinophils and IL13 originating from innate lymphoid type 2 cells and invariant natural killer T (iNKT) cells (Wu et al. 2011). Obesity leads to decreased expression of these factors while simultaneously increasing the expression of proinflammatory antigens such as F4/80, CD11b (integrin alpha M), and CD11c (integrin alpha X) and cytokines including TNFa, IL6, and nitric oxide synthase 2 (NOS2) resulting in a shift from the antiinflammatory M2 to proinflammatory M1 phenotype (Lumeng et al. 2007b). This shift is not induced by the transformation of resident M2 macrophages, but rather by increased recruitment of circulating monocytes and their differentiation into M1 cells as more than 90% of recruited monocytes become CD11c+ ATMs (Nguyen et al. 2007). Furthermore, this process requires a functioning chemotactic MCP1/CCR2 axis (Lumeng et al. 2007a). In addition to phenotypic changes, obesity alters also the morphology and localization of ATMs. Thus, unlike the M2 macrophages interspersed in the stromal-vascular fraction of AT newly recruited, M1 ATMs aggregate in specific clusters termed crown-like structures surrounding large lipid droplet remains of necrotic adipocytes and forming foam cells by accumulating lipids (Prieur et al. 2011).

There is ample evidence that the polarization state of ATMs significantly affects systemic inflammation and insulin action. The number of CD11c+ cells was shown to correlate with IR and their ablation ameliorated IR and reduced local as well as systemic production of

proinflammatory factors (Patsouris et al. 2008, Fujisaka et al. 2009). The potential mechanisms involved in these processes range from TNFa-mediated inhibition of insulin signaling and downregulation of GLUT4 transporter in adipocytes and increased production of collagen and fibrotic remodelation of extracellular matrix, to the recruitment and activation of other immune cells via secretion of chemokines and presentation of antigens and stimulatory signals (Lumeng et al. 2007c, Patsouris et al. 2008, Khan et al. 2009, Sun et al. 2011). Several candidate factors responsible for driving the M1 shift in ATM polarization have been identified so far, including toll-like receptors (TLRs), metabolic endotoxemia, lipid spillover, and adipokines. TLRs are pattern recognition receptors that activate innate immune responses by identifying foreign pathogens. Their ligands include various infectious antigens, among them bacterial lipopolysaccahrides (LPS) that bind to TLR4. Interestingly, obese mice showed increased LPS circulating levels as a result of enhanced LPS translocation from the gut and TLR-deficient mice exhibited decreased ATM numbers and reduced M1 polarization (Cani et al. 2007, Saberi et al. 2009). In humans, systemic LPS positively correlated with AT inflammation and IR (Creely et al. 2007). Importantly, other TLR4 ligands capable of activating antibacterial inflammatory response include saturated free fatty acids (FFA), heat shock proteins, and other substances elevated in obesity and T2DM (Dasu et al. 2010). Lipid spillover caused by chronically increased food intake and subsequent inability of adipocytes to store excess energy induce M1 macrophage shift also via the ER stress and inflammasome activation (Erbay et al. 2009, Vandanmagsar et al. 2011). In contrast to saturated FFAs, unsaturated fatty acids drive the shift toward M2 cells by binding to peroxisome proliferator-activated receptor γ (PPAR γ). The same mechanism seems to be partially responsible for the favorable antidiabetic and metabolic effects of other PPARy ligands - glitazones (Odegaard et al. 2007, Stienstra et al. 2008). The metabolically positive adipokine adiponectin also shows M2-polarizing effects; its decline with growing obesity might thus be one of the primary factors responsible for the M1 shift in ATMs (Ohashi et al. 2010).

Although instructive *in vitro*, a strict polarization to either M1 or M2 phenotype does not seem to capture the whole *in vivo* reality, especially in humans, as human (also rodent in some studies) ATMs were found to simultaneously express M1 (F4/80 and CD11c) as well as M2 (CD206 and CD301) markers (Bourlier *et al.* 2008, Shaul *et al.* 2010). Moreover, with increased BMI, human ATMs show decreased expression of several M1 markers while increasing the expression of M2 marker lymphatic vessel endothelial hyaluronan receptor 1 (LYVE1) and they are at the same time capable of producing proinflammatory as well as antiinflammatory factors (Zeyda *et al.* 2007). It was further demonstrated that the ATM phenotype is not fixed and that macrophages can repolarize from one state to another either by switching from high-fat diet to normal chow or by administering ω -3 FFA or glitazones, or increasing the levels of adiponectin (Bouhlel *et al.* 2007, Li *et al.* 2010, Oh *et al.* 2010). Thus, it seems that the M1/M2 classification is oversimplified and that a continuum rather than two distinctly opposite states exists between both phenotypes where the final effect results from the interaction of currently acting pro- and antiinflammatory stimuli.

Dendritic cells

Although macrophages, except of producing large quantities of cytokines and chemokines, are also capable of presenting antigens to effector lymphoid cells, it is their closest congeners, the DCs, that function as primary APCs of the immune system enabling the transition from innate to adaptive immunity by presenting antigens via major histocompatibility complex II (MHCII) molecules to the T cell receptors (TCRs) of CD4 helper T (Th) cells (Steinman 2008). In addition, DCs also produce an array of cytokines involved in the maturation and activation of adaptive immunity cells including IL12 that induces the differentiation of naïve T cells into Th1 phenotype and IL15 that helps in the proliferation of CD8⁺T lymphocytes and NK cells (Lee & Lee 2014).

DCs are probably the least explored subset of AT immune cells, partially due to their central role in adaptive immunity, whereas AT inflammation is considered primarily an innate immune response. Moreover, their identification in AT is complicated by the fact that one of their chief antigens, CD11c, is abundantly expressed also on proinflammatory M1 ATMs (in flow cytometry, ATMs are usually defined as F4/80⁺CD11b⁺CD11c⁺ cells while DCs are F4/80^{-/low}CD11b⁻CD11c⁺) (Dominguez & Ardavin 2010, Hashimoto et al. 2011). However, the recently suggested role of various types of T lymphocytes in triggering ATM recruitment turned the attention also to DCs as the main players in T cell differentiation. Indeed, it was shown that high-fat diet increases the AT DC content in murine models of obesity and that the expression of DC antigens CD1c, CD11c, and CD83 is elevated in the subcutaneous AT of obese humans as compared with their lean counterparts (Bertola et al. 2012). Moreover, DCs

lournal of Endocrinology

from obese subjects were *in vitro* able to induce the differentiation of Th17 cells (Th cells expressing IL17), while the amount of CD103 + DCs important for the differentiation of Treg cells was decreases (Bertola *et al.* 2012). The resulting imbalance between anti-inflammatory Treg and proinflammatory Th17 cells can lead to a Th1 shift in T-helper phenotype and subsequent M1 polarization of ATMs. In another study, genetic ablation of DCs was associated with decreased number of ATMs and liver macrophages and improved IR, although it is not clear whether these effects could be attributable solely to DC deletion or rather to the loss of weight that accompanied the mutation (Stefanovic-Racic *et al.* 2012).

Mast cells

Mast cells, abundant in barriers, such as skin and the mucosa, function as first-line responders to invading pathogens, mainly due to their rapid degranulation ability. Although their uncontrolled activation substantially contributes to the development of asthma, allergy, and anaphylaxis, recently it has been suggested that mast cells are also directly involved in defense reactions against bacterial and parasitic infections (Galli et al. 2005, Abraham & St John 2010). Activated mast cells secrete a broad spectrum of inflammatory mediators including histamine, heparin, lipid mediators (PGE₂ and LTB4), proteases (chymases and tryptases), and pro- and antiinflammatory cytokines (TNFa, IL1β, IL6, TGFβ, IL4, and IL10) (Abraham & St John 2010). Although not as abundant as macrophages, mast cell numbers are significantly elevated in the AT of obese mice and humans (Liu et al. 2009). Mast cell-deficient KitW-sh/KitW-sh mice or mice treated with a mast cell stabilizer (disodium cromoglycate) show almost no ATM infiltration along with improved insulin sensitivity and reduced body weight under the diet-induced obesity conditions, while mast cell reconstitution is accompanied by increased IR (Liu et al. 2009). Interestingly, IL6 and IFNy seem to play an important role in this process, as reconstitution with mast cells from Il6 or Ifng knockout mice had no effect on insulin sensitivity. The deficit in mast cells resulted also in decreased angiogenesis leading to the presumption that mast cells might regulate AT inflammation through vessel growth (Liu et al. 2009). Although certainly requiring further proof, this hypothesis might at least partially explain the changes in body weight and adiposity associated with mast cell modulation. Taken together, mast cells appear to modulate IR indirectly by influencing body weight and adiposity rather than by directly regulating AT inflammation.

Neutrophils

Neutrophils, along with eosinophils and basophils, belong to the granulocyte subgroup of myeloid immune cells, containing in their cytoplasm large numbers of granules with diverse biologically active substances including in case of neutrophils potent antibacterial agents as lysozyme, neutrophil elastase (NE), and myeloperoxidse (MPO). Neutrophils are considered as the primary effectors of acute inflammatory reaction as they are the first cells to be recruited to the site of inflammation where they fight the invading pathogens by degranulation of their antimicrobial reagents as well as by phagocytosis. Moreover, at the local infection site, neutrophils are capable of producing large quantities of cytokines and chemokines including TNF α , IL1 β , IL8, and CCL3, inducing thus the recruitment and activation of the second wave of immune cells, most notably macrophages, DCs, and lymphocytes (Mantovani et al. 2011, Amulic et al. 2012).

Their pivotal position in the initiation of inflammatory reactions raises the question about a possible role of neutrophils in AT inflammation. In obese individuals, plasma concentrations of MPO and calprotectin (a factor mainly derived from neutrophils) as well as the levels of neutrophil activation marker CD66b were increased compared with lean controls suggesting that obesity affects systemic activation of neutrophils (Nijhuis et al. 2009). Neutrophils were also found in the AT of lean mice, although they represent only a small fraction (<1%) of all AT immune cells (Ferrante 2013). Intriguingly, high-fat diet lead to a 20-fold increase in AT neutrophil (ATN) content occurring as early as 3 days after its initiation (in contrast to 7 days for macrophages (Nguyen et al. 2007)) thus making neutrophils the earliest immune cells to be recruited into AT. However, the MPO expression levels used to detect neutrophils decreased after the first weak, suggesting only a transient character of ATN infiltration (Elgazar-Carmon et al. 2008). Another study using CD11b⁺Ly6g⁺F4/80⁻CD11c⁻ cells was assessed by flow cytometry as ATNs also showed a rapid increase in ATNs after HFD, further strengthening the potential role of neutrophils in the initiation of AT inflammation. Conversely to previous results, this increase was sustained also after 90 days of high-fat feeding, indicating possible different roles of ATNs at different stages of obesity development (Talukdar et al. 2012). Furthermore, NE was found to be of significant importance for ATN recruitment and inflammation, as its genetic deletion or pharmacological inhibition suppressed AT inflammation (mainly by reduction of M1 macrophage numbers) and improved

http://joe.endocrinology-journals.org DOI: 10.1530/JOE-14-0283

obesity-related IR. Suggested mechanisms explaining this effect include the degradation of insulin receptor substrate 1 (IRS1) or the activation of TLR4 pathway by NE (Talukdar *et al.* 2012). All these results still await their confirmation in humans; however, if neutrophil recruitment is the initial event of AT inflammation, the identification of precise chemoattracting signals might open up novel possibilities for the treatment and prevention of obesity-related complications.

Eosinophils

Eosinophils are the primary effector cells in the defense against parasitic infections and they play a central role in the development of allergic reactions (Rosenberg *et al.* 2013). Unlike neutrophils that are involved in the antibacterial Th1 reactions, eosinophils are important mediators of Th2 immunity, producing a vast array of Th2 cytokines (e.g. IL4, IL10, IL13, and TGFB) that participate in anti-inflammatory immune responses, M2 polarization of macrophages, and differentiation of Th2 cells (Spencer & Weller 2010).

IL4 was shown to have insulin-sensitizing effects, as deletion of Stat6, a signaling molecule instrumental for mediating the effects of IL4, decreases insulin sensitivity, while systemic infusion of IL4 leads to amelioration of IR (Ricardo-Gonzalez et al. 2010). Despite being present at very low numbers (cca 20 000 eosinophils/g of fat), using Il4 reporter mice it was demonstrated that 90% of ATproduced IL4 originates from resident eosinophils (Wu et al. 2011). As IL4 and IL13 are considered as main mediators of M2 polarization of ATMs, it was hypothesized that eosinophils might be the central driver of M2 differentiation. Indeed, Wu et al. (2011) reported that ATresident eosinophil numbers correlate positively with M2 ATMs and that M2 polarization is mediated by eosinophils in an IL4/IL13-dependent fashion. Furthermore, obesity decreased AT eosinophil numbers leading to reduced insulin sensitivity, while the increase in eosinophils due to the overexpression of IL5 or helminth infection improved obesity-induced IR (Wu et al. 2011). However, as body weight changes occurred in all of the studied mice models, it is currently not clear, whether the eosinophilmediated regulation of obesity-induced IR and AT inflammation can be attributed to the direct effects of eosinophils on IR or whether it is caused by secondary effects of eosinophils on changes in body weight and adiposity. Recently, it has been also reported that AT eosinophils themselves depend upon the IL5- and IL13-producing innate lymphoid type 2 cells (Molofsky et al. 2013).

All in all, the eosinophil studies indicate the existence of a new pathway that is able to improve AT inflammation and obesity-related IR by inducing Th2 immune response that either modulates AT inflammation directly or indirectly via changes in adiposity.

AT lymphoid cells

As already mentioned, the lymphoid line consists of T- and B lymphocytes, NK cells, and NKT cells, all of which are produced in the bone marrow. By recognizing specific antigens with their receptors, T- and B lymphocytes play important roles in adaptive immunity. In contrast, NK and NKT cells are supposed to be involved more in innate immunity, although the new data suggest their significance also in adaptive immunity (Kondo *et al.* 2003, Lee & Lee 2014).

T cells

T cells are bone marrow-derived lymphocytes that fully mature in the thymus (Koch & Radtke 2011). They play a major role in adaptive immunity by shifting from naive to several effector states during an immune response (Jager & Kuchroo 2010). Based on the expression of surface markers, T cells can be divided into CD4⁺ and CD8⁺ subtypes and according to their function into Th cells, cytotoxic T cells, regulatory T cells, and others. Most of Th cells express CD4 and can be further categorized according to the production of specific cytokines into Th1 (signature cytokine IFN_Y), Th2 (signature cytokines IL4, IL5, and IL13), Th17 (signature cytokines IL17, IL21, and IL22), and Treg (signature cytokines IL10 and transforming growth factor β – TGFB) (Oestreich & Weinmann 2012). In contrast, CD8⁺T cells are considered mainly cytotoxic, even though, depending on the conditions, CD4⁺ cells can also exert cytotoxic activity (Zhang & Bevan 2011). CD8⁺ cells produce a variety of cytolytic substances including perforins and granzymes, but also secrete a number of cytokines that regulate the development and activation of other immune cells (Lee & Lee 2014). In AT, T cells (CD3+) constitute the second largest immune cell population in AT after ATMs and obesity along with increasing their total numbers also alters the proportions of different T cell subsets. The resulting derangements in adaptive as well as innate immune reactions seem to play important roles in the development of obesity-related inflammation, mainly by influencing AMT numbers and activation state.

http://joe.endocrinology-journals.org DOI: 10.1530/JOE-14-0283

CD4⁺T cells

CD4⁺ Th1 cells were shown to be increased in obesity and their density correlated positively with the incidence of nonalcoholic fatty liver disease (Pacifico et al. 2006). Moreover, their main secretory product, IFN γ , seems to promote M1 polarization of ATMs, as IFNy-deficient mice exert reduced AT inflammation and ameliorated IR without changes in body weight (Rocha et al. 2008). Similar results were obtained when modulating IFN γ CD4⁺ cells into regulatory T cells (Winer et al. 2009). As mentioned in connection with mast cells, the role of IFN γ in AT inflammation is still not completely understood, but it most probably involves the modulation of oxidative metabolism and microangiogenesis to facilitate macrophage infiltration (Wong et al. 2011). Nevertheless, these data indicate a significant role for Th1 cells and IFN γ in the mediation of AT inflammation and obesity-related IR.

The complex effect of T cells on inflammatory processes in AT is best demonstrated by the results of Winer *et al*. who found that in $Rag^{-/-}$ mice with congenital deficit of T-and B cells (RAGs - recombination-activating genes - are vital for the recombination of TCRs in T lymphocytes and immunoglobulins in B lymphocytes and their knockout abolishes both lymphocyte populations), high-fat diet induced a greater degree of IR compared with WT controls. Reconstruction of CD4 T cells decreased their body weight and improved insulin sensitivity; however, when using CD4 T cells from Stat6-/knockout mice (which have impaired development of Th2 cells but normal development of Th1 lymphocytes) no improvement could be seen at all (Winer et al. 2009). This suggests that the anti-inflammatory CD4⁺ Th2 cells play a suppressive role in the development of obesity-related inflammation and IR, and the shift in Th1/Th2 ratio toward the proinflammatory Th1 phenotype might be responsible for the polarization from M2 to M1 ATMs.

Interestingly, adipose $CD4^+T$ cells show only a limited TCR repertoire, which is different from T cell populations in other tissues (e.g. spleen), arguing that a specific set of antigens drives the polarization of T cells in AT (Winer *et al.* 2009). Although the existence of such antigens remains to be confirmed, their identification might potentially offer new therapeutic targets for modulation of early phases of AT inflammation.

CD8⁺T cells

CD8⁺T cells as the chief cytotoxic immune cells are involved mainly in the antiviral response, producing

http://joe.endocrinology-journals.org DOI: 10.1530/JOE-14-0283 © 2014 Society for Endocrinology Printed in Great Britain increases the numbers of CD8⁺T lymphocytes (three- to four-times as compared with lean state) along with increased expression of their products, most notably granzyme B and IFNy. Furthermore, CD8⁺T cell infiltration precedes the infiltration of macrophages into AT, and CD8⁺T lymphocytes stimulate M1 macrophage polarization in vitro as well as in vivo (Rausch et al. 2008). It was also shown that depletion of CD8⁺T cells in obese rodents improved insulin sensitivity, while their adoptive transfer into CD8-deficient animals lead to increased M1 ATM accumulation in AT and to the development of IR (Nishimura et al. 2009). Thus, CD8⁺T cells also seem to be involved in early phases of ATM recruitment and M1 polarization. Surprisingly, CD8⁺T cell deficit does not fully prevent the insulin-resistant phenotype when challenged with high-fat diet as well as adoptive transfer of CD8⁺ cells into lymphocyte of naïve $Rag^{-/-}$ mice does not further aggravate the preexisting IR, suggesting that other factors or immune cells might be required to mediate the full effect of CD8⁺T cells (Nishimura et al. 2009, Winer et al. 2009).

cytolytic molecules upon activation by MHCI antigens on APCs (Sun *et al.* 2012). As with CD4⁺ cells, obesity also

Th17 cells

Th17 cells are involved in autoimmune disorders along producing Th17-specific cytokines, IL17 and IL23, which are thought to initiate pathogenic inflammation. In obese subjects with and without T2DM, increased serum concentrations of these Th17 signature cytokines along with elevated numbers of Th17 cells and decreased amount of the antagonist Treg lymphocytes could be found (Zuniga et al. 2010, Jagannathan-Bogdan et al. 2011, Goossens et al. 2012). IL17 might induce IR by activating JNK, which in turn interferes with insulin receptor signaling on the level of IRS1 (Zhu et al. 2011). IL17 is also implicated to be involved in the development of atherosclerosis and cardiovascular diseases (Ding et al. 2012). In contrast, Zuniga et al. (2010) suggested that IL17 has protective effects against obesity and IR as IL17 deficiency enhanced AT accumulation and increased fasting glucose even in mice on low-fat diet. To address these contradictions, the local and systemic effects of IL17 will certainly require further investigation.

Recently, $\gamma \delta T$ cells, which are T lymphocytes with $\gamma \delta T$ receptor (in contrast to standard $\alpha\beta TRC$) that have only a restricted antigen repertoire and are unable to develop immunological memory (and as such stand at the cross-road between innate and adaptive immunity), have been

Published by Bioscientifica Ltd.

identified as the main source of IL17 in AT (Caspar-Bauguil *et al.* 2005, Zuniga *et al.* 2010). As the secretion of IL17 is induced by IL1 β , which by itself is produced as a result of lipid-mediated inflammasome activation, lipid spillover might be the main initiator of IL17 proinflammatory reaction in AT.

T regulatory cells

CD4⁺CD25+Foxp3+ regulatory T cells (Treg) are considered as suppressors of inflammatory reactions as they are vital for maintaining self-tolerance and curbing the proinflammatory Th1 and Th17 responses. Two distinct subsets of Treg can be distinguished, natural Treg (nTreg) and induced Treg (iTreg), which differentiate from the naïve T cells under the influence of IL2 and TGFB (Kretschmer et al. 2005). Under lean conditions, IL10 produced by Tregs helps in preserving an anti-inflammatory environment. In contrast to Th1 and cytotoxic T cells, obesity decreases the number of Tregs in AT (Feuerer et al. 2009). AT Treg depletion increases IR as well as local and systemic production of proinflammatory cytokines, whereas exogenous IL2-mediated stimulation of Treg population is accompanied by increase in IL10 levels and amelioration of IR (Feuerer et al. 2009). Several mechanisms by which Tregs might improve AT inflammation and obesity-related IR have been suggested including increased glucose uptake into adipocytes, reduction in ATM M1 polarization (as the number of Tregs inversely correlates with M1 macrophages in AT), and prevention of Th1 differentiation of AT-resident T cells (Feuerer et al. 2009, Deiuliis et al. 2011). Moreover, it was shown that AT Tregs had markedly increased the expression of $PPAR\gamma$ (master regulator of adipocyte differentiation) as compared with Tregs from other tissues and that treatment with $PPAR\gamma$ agonists thiazolidinediones (TZDs) resulted in elevated Treg numbers in AT (Cipolletta et al. 2012). Interestingly, Treg-specific deletion of PPARy reduced Tregs only in AT but not in spleen and TZD treatment had no effect on obesity-induced IR in these mice, whereas in their WT littermates TZDs improved blood glucose and insulin sensitivity along with the increase in AT Treg numbers (Cipolletta et al. 2012). Thus, it appears that TZDs exert their favorable metabolic effects at least partially via targeting AT Tregs.

NKT cells

NKT cells are a highly specialized T cell subpopulation with potent immunomodulatory functions, which instead

http://joe.endocrinology-journals.org DOI: 10.1530/JOE-14-0283 of peptide antigens presented by MHCI or MHCII molecules respond to lipid antigens presented mainly by the antigen-presenting molecule CD1d (Kronenberg 2005, Bendelac *et al.* 2007, Wu *et al.* 2012). NKT cells can be categorized into two basic subgroups according to the variation in their T cell receptor (TCR) sequence: type I or invariant NKT (iNKT) cells with in variant α -chain of TCR and type II or variant NKT (vNKT) cells with a more diverse TCR sequence (Godfrey *et al.* 2004). NKT cells secrete a number of different cytokines including proinflammatory TNF α and IFN γ as well as anti-inflammatory IL4 and IL13 and can be thus involved in both Th1 and Th2 responses (Lee & Lee 2014).

NKT cells form the major part of liver T cells (30-50%) and their deletion leads to hepatic steatosis. In lean subjects, iNKT cells seem to be enriched in AT than in systemic circulation (Lynch et al. 2009). Under obese condition, the number of iNKT cells is reduced in AT as well as in peripheral circulation and liver (Lynch et al. 2012). However, the exact role of iNKT cells in obesityrelated inflammation and IR is still unclear, as studies on iNKT cell-depleted mice on high-fat diet yielded contradictory results ranging from improvement (Ohmura et al. 2010, Satoh et al. 2012, Wu et al. 2012) to no effect (Kotas et al. 2011, Mantell et al. 2011) and to worsening (Lynch et al. 2012, Schipper et al. 2012b) of AT inflammation and insulin sensitivity as compared with WT controls. Decreased AT iNKT numbers in obesity might point to a potential role of iNKT cells in the maintenance of antiinflammatory AT profile in lean individuals; nevertheless further research is clearly needed to dissect the significance of iNKT cells in immunometabolic reactions.

B cells

B cells are unique immune cells that emerge from the bone marrow in immature form and then fully mature in the secondary lymphoid organs (spleen and lymph nodes). Their chief function is the promotion of humoral immunity by producing antibodies specific for foreign antigens, but they can also act as APCs via MHCI and MHCII molecules. B cells are also capable of recognizing certain pathogen-associated patterns via specific TLRs (LeBien & Tedder 2008). Obesity increases the number of B cells in AT (most notably IgG-producing cells), while B cell depletion was shown to improve IR (Winer *et al.* 2011, DeFuria *et al.* 2013). Moreover, B cell-derived IgG2c antibodies are elevated in obese animals and their transfer into lean mice resulted in AT inflammation and development of IR. This reaction required the presence of T cells,

lournal of Endocrinology

most notably CD4⁺ and/or CD8⁺ cells (Winer *et al.* 2011). As IgG2c antibodies were preferentially found in crownlike structures, indicating their role in the clearance of necrotic adipocytes. As the recruitment of B cells into AT preceded M1 polarization of ATMs, IgG2c antibodies were suggested to drive macrophage polarization toward the M1 phenotype (Duffaut *et al.* 2009). This hypothesis was further strengthened by the fact that B cell-deficient mice exert reduced AT M1 polarization, while IgG2c induce TNF α production in macrophages *in vitro* (Winer *et al.* 2011). Taken together, the finding that a B cell-produced set of antibodies can play a direct role in the pathogenesis of T2DM might potentially change the paradigm of obesity-induced inflammation as an innate immune response and therefore requires further confirmation.

Other lymphoid cells

Recently, innate lymphoid type 2 cells, which are CD25, IL7, IL33, and GATA-binding protein 3 positive, have been identified in lean AT. As these cells are the predominant producers of AT IL5 and IL13, they might be crucial for maintaining the M2 ATM- and eosinophil-rich AT profile associated with good metabolic health and the absence of AT inflammation (Molofsky *et al.* 2013).

Conclusion and future directions

In recent years, AT has emerged as a highly active organ integrating metabolic, endocrine, and immune functions into a single entity that exerts significant effects on wholebody homeostasis. Under physiological conditions, the structural and functional integrity of AT are sustained by a meticulously orchestrated network of immune cells and reactions. However, due to the proximity of metabolic and immune pathways, chronic overnutrition induces severe derangements in this network by changing the amount and activity of almost all resident immune cells and promoting the recruitment of different immune cell subsets. The resulting imbalance in immunological phenotypes leads to the development of local inflammation that by releasing biologically active substances further spreads into systemic circulation thus affecting also diverse remote organs. Although much work has been done in elucidating the mechanisms by which particular immune cells contribute to AT inflammation, an even greater number of questions remain open, most notably concerning the precise character and sequence of insults that initiate the inflammatory response. Nevertheless, the inflammatory nature of obesity opens up new horizons in the development of obesity-related treatment strategies including targeted modulation of key elements and processes responsible for the transition from simple adiposity to subsequent metabolic, cardiovascular, and other complications.

Declaration of interest

The authors declare that there is no conflict of interest that could be perceived as prejudicing the impartiality of the review.

Funding

This work was supported by grants from Czech Republic Ministry of Health (RVO-VFN64165 and IGA NT13299-4).

References

- Abraham SN & St John AL 2010 Mast cell-orchestrated immunity to pathogens. *Nature Reviews. Immunology* **10** 440–452. (doi:10.1038/ nri2782)
- Al-Lahham S, Roelofsen H, Rezaee F, Weening D, Hoek A, Vonk R & Venema K 2012 Propionic acid affects immune status and metabolism in adipose tissue from overweight subjects. *European Journal of Clinical Investigation* 42 357–364. (doi:10.1111/j.1365-2362.2011.02590.x)
- Amulic B, Cazalet C, Hayes GL, Metzler KD & Zychlinsky A 2012 Neutrophil function: from mechanisms to disease. *Annual Review of Immunology* **30** 459–489. (doi:10.1146/annurev-immunol-020711-074942)
- Apovian CM, Bigornia S, Mott M, Meyers MR, Ulloor J, Gagua M, McDonnell M, Hess D, Joseph L & Gokce N 2008 Adipose macrophage infiltration is associated with insulin resistance and vascular endothelial dysfunction in obese subjects. *Arteriosclerosis, Thrombosis, and Vascular Biology* **28** 1654–1659. (doi:10.1161/ATVBAHA.108.170316)
- Bendelac A, Savage PB & Teyton L 2007 The biology of NKT cells. *Annual Review of Immunology* **25** 297–336. (doi:10.1146/annurev.immunol.25. 022106.141711)
- Bertola A, Ciucci T, Rousseau D, Bourlier V, Duffaut C, Bonnafous S, Blin-Wakkach C, Anty R, Iannelli A, Gugenheim J *et al.* 2012 Identification of adipose tissue dendritic cells correlated with obesityassociated insulin-resistance and inducing Th17 responses in mice and patients. *Diabetes* **61** 2238–2247. (doi:10.2337/db11-1274)
- Bluher M 2009 Adipose tissue dysfunction in obesity. *Experimental and Clinical Endocrinology & Diabetes* **117** 241–250. (doi:10.1055/s-0029-1192044)
- Bouhlel MA, Derudas B, Rigamonti E, Dievart R, Brozek J, Haulon S, Zawadzki C, Jude B, Torpier G, Marx N *et al.* 2007 PPARγ activation primes human monocytes into alternative M2 macrophages with anti-inflammatory properties. *Cell Metabolism* **6** 137–143. (doi:10.1016/ j.cmet.2007.06.010)
- Bourlier V, Zakaroff-Girard A, Miranville A, De Barros S, Maumus M, Sengenes C, Galitzky J, Lafontan M, Karpe F, Frayn KN *et al.* 2008 Remodeling phenotype of human subcutaneous adipose tissue macrophages. *Circulation* **117** 806–815. (doi:10.1161/CIRCULATION-AHA.107.724096)
- Burcelin R, Serino M, Chabo C, Garidou L, Pomie C, Courtney M, Amar J & Bouloumie A 2013 Metagenome and metabolism: the tissue microbiota hypothesis. *Diabetes, Obesity & Metabolism* **15**(Suppl 3) 61–70. (doi:10.1111/dom.12157)
- Cancello R, Henegar C, Viguerie N, Taleb S, Poitou C, Rouault C, Coupaye M, Pelloux V, Hugol D, Bouillot JL *et al.* 2005 Reduction of

http://joe.endocrinology-journals.org DOI: 10.1530/JOE-14-0283 Published by Bioscientifica Ltd.

macrophage infiltration and chemoattractant gene expression changes in white adipose tissue of morbidly obese subjects after surgery-induced weight loss. *Diabetes* **54** 2277–2286. (doi:10.2337/diabetes.54.8.2277)

- Cancello R, Tordjman J, Poitou C, Guilhem G, Bouillot JL, Hugol D, Coussieu C, Basdevant A, Bar Hen A, Bedossa P *et al.* 2006 Increased infiltration of macrophages in omental adipose tissue is associated with marked hepatic lesions in morbid human obesity. *Diabetes* **55** 1554–1561. (doi:10.2337/db06-0133)
- Cani PD, Amar J, Iglesias MA, Poggi M, Knauf C, Bastelica D, Neyrinck AM, Fava F, Tuohy KM, Chabo C *et al.* 2007 Metabolic endotoxemia initiates obesity and insulin resistance. *Diabetes* **56** 1761–1772. (doi:10.2337/ db06-1491)
- Carvalho BM & Saad MJ 2013 Influence of gut microbiota on subclinical inflammation and insulin resistance. *Mediators of Inflammation* **2013** 986734. (doi:10.1155/2013/986734)
- Caspar-Bauguil S, Cousin B, Galinier A, Segafredo C, Nibbelink M, Andre M, Casteilla L & Penicaud L 2005 Adipose tissues as an ancestral immune organ: site-specific change in obesity. *FEBS Letters* **579** 3487–3492. (doi:10.1016/j.febslet.2005.05.031)
- Charriere G, Cousin B, Arnaud E, Andre M, Bacou F, Penicaud L & Casteilla L 2003 Preadipocyte conversion to macrophage. Evidence of plasticity. *Journal of Biological Chemistry* 278 9850–9855. (doi:10.1074/jbc. M210811200)
- Chawla A, Nguyen KD & Goh YP 2011 Macrophage-mediated inflammation in metabolic disease. *Nature Reviews. Immunology* **11** 738–749. (doi:10.1038/nri3071)
- Chen A, Mumick S, Zhang C, Lamb J, Dai H, Weingarth D, Mudgett J, Chen H, MacNeil DJ, Reitman ML *et al.* 2005 Diet induction of monocyte chemoattractant protein-1 and its impact on obesity. *Obesity Research* **13** 1311–1320. (doi:10.1038/oby.2005.159)
- Christiansen T, Richelsen B & Bruun JM 2005 Monocyte chemoattractant protein-1 is produced in isolated adipocytes, associated with adiposity and reduced after weight loss in morbid obese subjects. *International Journal of Obesity* **29** 146–150. (doi:10.1038/sj.ijo.0802839)
- Cildir G, Akincilar SC & Tergaonkar V 2013 Chronic adipose tissue inflammation: all immune cells on the stage. *Trends in Molecular Medicine* **19** 487–500. (doi:10.1016/j.molmed.2013.05.001)
- Cipolletta D, Feuerer M, Li A, Kamei N, Lee J, Shoelson SE, Benoist C & Mathis D 2012 PPAR-γ is a major driver of the accumulation and phenotype of adipose tissue Treg cells. *Nature* **486** 549–553. (doi:10.1038/nature11132)
- Cox LM & Blaser MJ 2013 Pathways in microbe-induced obesity. *Cell Metabolism* 17 883–894. (doi:10.1016/j.cmet.2013.05.004)
- Creely SJ, McTernan PG, Kusminski CM, Fisher FM, Da Silva NF, Khanolkar M, Evans M, Harte AL & Kumar S 2007 Lipopolysaccharide activates an innate immune system response in human adipose tissue in obesity and type 2 diabetes. *American Journal of Physiology. Endocrinology and Metabolism* **292** E740–E747. (doi:10.1152/ajpendo. 00302.2006)
- Curat CA, Miranville A, Sengenes C, Diehl M, Tonus C, Busse R & Bouloumie A 2004 From blood monocytes to adipose tissue-resident macrophages: induction of diapedesis by human mature adipocytes. *Diabetes* **53** 1285–1292. (doi:10.2337/diabetes.53.5.1285)
- Curat CA, Wegner V, Sengenes C, Miranville A, Tonus C, Busse R & Bouloumie A 2006 Macrophages in human visceral adipose tissue: increased accumulation in obesity and a source of resistin and visfatin. *Diabetologia* **49** 744–747. (doi:10.1007/s00125-006-0173-z)
- Dasu MR, Devaraj S, Park S & Jialal I 2010 Increased toll-like receptor (TLR) activation and TLR ligands in recently diagnosed type 2 diabetic subjects. *Diabetes Care* **33** 861–868. (doi:10.2337/dc09-1799)
- DeFuria J, Belkina AC, Jagannathan-Bogdan M, Snyder-Cappione J, Carr JD, Nersesova YR, Markham D, Strissel KJ, Watkins AA, Zhu M *et al.* 2013 B cells promote inflammation in obesity and type 2 diabetes through regulation of T cell function and an inflammatory cytokine profile. *PNAS* **110** 5133–5138. (doi:10.1073/pnas.1215840110)

Deiuliis J, Shah Z, Shah N, Needleman B, Mikami D, Narula V, Perry K, Hazey J, Kampfrath T, Kollengode M *et al.* 2011 Visceral adipose inflammation in obesity is associated with critical alterations in tregulatory cell numbers. *PLoS ONE* **6** e16376. (doi:10.1371/journal. pone.0016376)

Ding HS, Yang J, Yang J, Ding JW, Chen P & Zhu P 2012 Interleukin-17 contributes to cardiovascular diseases. *Molecular Biology Reports* **39** 7473–7478. (doi:10.1007/s11033-012-1580-5)

Dolezalova R, Lacinova Z, Dolinkova M, Kleiblova P, Haluzikova D, Housa D, Papezova H & Haluzik M 2007 Changes of endocrine function of adipose tissue in anorexia nervosa: comparison of circulating levels versus subcutaneous mRNA expression. *Clinical Endocrinology* **67** 674–678. (doi:10.1111/j.1365-2265.2007.02944.x)

- Dominguez PM & Ardavin C 2010 Differentiation and function of mouse monocyte-derived dendritic cells in steady state and inflammation. *Immunological Reviews* 234 90–104. (doi:10.1111/j.0105-2896.2009. 00876.x)
- Duffaut C, Galitzky J, Lafontan M & Bouloumie A 2009 Unexpected trafficking of immune cells within the adipose tissue during the onset of obesity. *Biochemical and Biophysical Research Communications* 384 482–485. (doi:10.1016/j.bbrc.2009.05.002)
- Elgazar-Carmon V, Rudich A, Hadad N & Levy R 2008 Neutrophils transiently infiltrate intra-abdominal fat early in the course of high-fat feeding. *Journal of Lipid Research* **49** 1894–1903. (doi:10.1194/jlr. M800132-JLR200)
- Erbay E, Babaev VR, Mayers JR, Makowski L, Charles KN, Snitow ME, Fazio S, Wiest MM, Watkins SM, Linton MF *et al.* 2009 Reducing endoplasmic reticulum stress through a macrophage lipid chaperone alleviates atherosclerosis. *Nature Medicine* **15** 1383–1391. (doi:10.1038/nm.2067)
- Feingold KR, Soued M, Staprans I, Gavin LA, Donahue ME, Huang BJ, Moser AH, Gulli R & Grunfeld C 1989 Effect of tumor necrosis factor (TNF) on lipid metabolism in the diabetic rat. Evidence that inhibition of adipose tissue lipoprotein lipase activity is not required for TNF-induced hyperlipidemia. *Journal of Clinical Investigation* 83 1116–1121. (doi:10.1172/JCI113991)
- Ferrante AW Jr 2013 The immune cells in adipose tissue. *Diabetes, Obesity & Metabolism* **15**(Suppl 3) 34–38. (doi:10.1111/dom.12154)
- Feuerer M, Herrero L, Cipolletta D, Naaz A, Wong J, Nayer A, Lee J, Goldfine AB, Benoist C, Shoelson S *et al.* 2009 Lean, but not obese, fat is enriched for a unique population of regulatory T cells that affect metabolic parameters. *Nature Medicine* **15** 930–939. (doi:10.1038/nm. 2002)
- Fujisaka S, Usui I, Bukhari A, Ikutani M, Oya T, Kanatani Y, Tsuneyama K, Nagai Y, Takatsu K, Urakaze M *et al.* 2009 Regulatory mechanisms for adipose tissue M1 and M2 macrophages in diet-induced obese mice. *Diabetes* 58 2574–2582. (doi:10.2337/db08-1475)

Galli SJ, Kalesnikoff J, Grimbaldeston MA, Piliponsky AM, Williams CM & Tsai M 2005 Mast cells as "tunable" effector and immunoregulatory cells: recent advances. *Annual Review of Immunology* **23** 749–786. (doi:10.1146/annurev.immunol.21.120601.141025)

Galli SJ, Borregaard N & Wynn TA 2011 Phenotypic and functional plasticity of cells of innate immunity: macrophages, mast cells and neutrophils. *Nature Immunology* **12** 1035–1044. (doi:10.1038/ni.2109)

Gerhardt CC, Romero IA, Cancello R, Camoin L & Strosberg AD 2001 Chemokines control fat accumulation and leptin secretion by cultured human adipocytes. *Molecular and Cellular Endocrinology* **175** 81–92. (doi:10.1016/S0303-7207(01)00394-X)

Godfrey DI, MacDonald HR, Kronenberg M, Smyth MJ & Van Kaer L 2004 NKT cells: what's in a name? *Nature Reviews. Immunology* **4** 231–237. (doi:10.1038/nri1309)

Goossens GH, Blaak EE, Theunissen R, Duijvestijn AM, Clement K, Tervaert JW & Thewissen MM 2012 Expression of NLRP3 inflammasome and T cell population markers in adipose tissue are associated with insulin resistance and impaired glucose metabolism in humans. *Molecular Immunology* **50** 142–149. (doi:10.1016/j.molimm.2012. 01.005) Journal of Endocrinology

222:3

- Gordon S 2003 Alternative activation of macrophages. *Nature Reviews*. *Immunology* **3** 23–35. (doi:10.1038/nri978)
- Harman-Boehm I, Bluher M, Redel H, Sion-Vardy N, Ovadia S, Avinoach E, Shai I, Kloting N, Stumvoll M, Bashan N *et al.* 2007 Macrophage infiltration into omental versus subcutaneous fat across different populations: effect of regional adiposity and the comorbidities of obesity. *Journal of Clinical Endocrinology and Metabolism* **92** 2240–2247. (doi:10.1210/jc.2006-1811)
- Hashimoto I, Wada J, Hida A, Baba M, Miyatake N, Eguchi J, Shikata K & Makino H 2006 Elevated serum monocyte chemoattractant protein-4 and chronic inflammation in overweight subjects. *Obesity* **14** 799–811. (doi:10.1038/oby.2006.93)
- Hashimoto D, Miller J & Merad M 2011 Dendritic cell and macrophage heterogeneity *in vivo. Immunity* **35** 323–335. (doi:10.1016/j.immuni. 2011.09.007)
- Heilbronn LK & Campbell LV 2008 Adipose tissue macrophages, low grade inflammation and insulin resistance in human obesity. *Current Pharmaceutical Design* **14** 1225–1230. (doi:10.2174/ 138161208784246153)
- Hirosumi J, Tuncman G, Chang L, Gorgun CZ, Uysal KT, Maeda K, Karin M & Hotamisligil GS 2002 A central role for JNK in obesity and insulin resistance. *Nature* **420** 333–336. (doi:10.1038/nature01137)
- Hotamisligil GS, Shargill NS & Spiegelman BM 1993 Adipose expression of tumor necrosis factor-α: direct role in obesity-linked insulin resistance. *Science* **259** 87–91. (doi:10.1126/science.7678183)
- Huber J, Kiefer FW, Zeyda M, Ludvik B, Silberhumer GR, Prager G, Zlabinger GJ & Stulnig TM 2008 CC chemokine and CC chemokine receptor profiles in visceral and subcutaneous adipose tissue are altered in human obesity. *Journal of Clinical Endocrinology and Metabolism* 93 3215–3221. (doi:10.1210/jc.2007-2630)
- Jagannathan-Bogdan M, McDonnell ME, Shin H, Rehman Q, Hasturk H, Apovian CM & Nikolajczyk BS 2011 Elevated proinflammatory cytokine production by a skewed T cell compartment requires monocytes and promotes inflammation in type 2 diabetes. *Journal of Immunology* **186** 1162–1172. (doi:10.4049/jimmunol.1002615)
- Jager A & Kuchroo VK 2010 Effector and regulatory T cell subsets in autoimmunity and tissue inflammation. *Scandinavian Journal of Immunology* 72 173–184. (doi:10.1111/j.1365-3083.2010.02432.x)
- Kanda H, Tateya S, Tamori Y, Kotani K, Hiasa K, Kitazawa R, Kitazawa S, Miyachi H, Maeda S, Egashira K *et al.* 2006 MCP-1 contributes to macrophage infiltration into adipose tissue, insulin resistance, and hepatic steatosis in obesity. *Journal of Clinical Investigation* **116** 1494–1505. (doi:10.1172/JCI26498)
- Khan T, Muise ES, Iyengar P, Wang ZV, Chandalia M, Abate N, Zhang BB, Bonaldo P, Chua S & Scherer PE 2009 Metabolic dysregulation and adipose tissue fibrosis: role of collagen VI. *Molecular and Cellular Biology* 29 1575–1591. (doi:10.1128/MCB.01300-08)
- Kim S, Kim JH, Park BO & Kwak YS 2014 Perspectives on the therapeutic potential of short-chain fatty acid receptors. *BMB Reports* **47** 173–178. (doi:10.5483/BMBRep.2014.47.3.272)
- Koch U & Radtke F 2011 Mechanisms of T cell development and transformation. Annual Review of Cell and Developmental Biology 27 539–562. (doi:10.1146/annurev-cellbio-092910-154008)
- Kondo M, Wagers AJ, Manz MG, Prohaska SS, Scherer DC, Beilhack GF, Shizuru JA & Weissman IL 2003 Biology of hematopoietic stem cells and progenitors: implications for clinical application. *Annual Review of Immunology* **21** 759–806. (doi:10.1146/annurev.immunol.21.120601. 141007)
- Kotas ME, Lee HY, Gillum MP, Annicelli C, Guigni BA, Shulman GI & Medzhitov R 2011 Impact of CD1d deficiency on metabolism. *PLoS* ONE 6 e25478. (doi:10.1371/journal.pone.0025478)
- Kremen J, Dolinkova M, Krajickova J, Blaha J, Anderlova K, Lacinova Z, Haluzikova D, Bosanska L, Vokurka M, Svacina S *et al.* 2006 Increased subcutaneous and epicardial adipose tissue production of proinflammatory cytokines in cardiac surgery patients: possible role in

postoperative insulin resistance. *Journal of Clinical Endocrinology and Metabolism* **91** 4620–4627. (doi:10.1210/jc.2006-1044)

- Kretschmer K, Apostolou I, Hawiger D, Khazaie K, Nussenzweig MC & von Boehmer H 2005 Inducing and expanding regulatory T cell populations by foreign antigen. *Nature Immunology* **6** 1219–1227. (doi:10.1038/ni1265)
- Kronenberg M 2005 Toward an understanding of NKT cell biology: progress and paradoxes. *Annual Review of Immunology* 23 877–900. (doi:10.1146/ annurev.immunol.23.021704.115742)
- LeBien TW & Tedder TF 2008 B lymphocytes: how they develop and function. *Blood* **112** 1570–1580. (doi:10.1182/blood-2008-02-078071)
- Leclerc V & Reichhart JM 2004 The immune response of *Drosophila* melanogaster. Immunological Reviews **198** 59–71. (doi:10.1111/j.0105-2896.2004.0130.x)
- Lee BC & Lee J 2014 Cellular and molecular players in adipose tissue inflammation in the development of obesity-induced insulin resistance. *Biochimica et Biophysica Acta* **1842** 446–462. (doi:10.1016/j. bbadis.2013.05.017)
- Li P, Lu M, Nguyen MT, Bae EJ, Chapman J, Feng D, Hawkins M, Pessin JE, Sears DD, Nguyen AK *et al.* 2010 Functional heterogeneity of CD11cpositive adipose tissue macrophages in diet-induced obese mice. *Journal of Biological Chemistry* 285 15333–15345. (doi:10.1074/jbc. M110.100263)
- Liu J, Divoux A, Sun J, Zhang J, Clement K, Glickman JN, Sukhova GK, Wolters PJ, Du J, Gorgun CZ *et al.* 2009 Genetic deficiency and pharmacological stabilization of mast cells reduce diet-induced obesity and diabetes in mice. *Nature Medicine* **15** 940–945. (doi:10.1038/ nm.1994)
- Lumeng CN, Bodzin JL & Saltiel AR 2007a Obesity induces a phenotypic switch in adipose tissue macrophage polarization. *Journal of Clinical Investigation* **117** 175–184. (doi:10.1172/JCI29881)
- Lumeng CN, Deyoung SM, Bodzin JL & Saltiel AR 2007b Increased inflammatory properties of adipose tissue macrophages recruited during diet-induced obesity. *Diabetes* 56 16–23. (doi:10.2337/ db06-1076)
- Lumeng CN, Deyoung SM & Saltiel AR 2007c Macrophages block insulin action in adipocytes by altering expression of signaling and glucose transport proteins. *American Journal of Physiology. Endocrinology and Metabolism* 292 E166–E174. (doi:10.1152/ajpendo.00284.2006)
- Lynch L, O'Shea D, Winter DC, Geoghegan J, Doherty DG & O'Farrelly C 2009 Invarient NKT cells and CD1d(+) cells amass in human omentum and are depleted in patients with cancer and obesity. *European Journal of Immunology* **39** 1893–1901. (doi:10.1002/eji.200939349)
- Lynch L, Nowak M, Varghese B, Clark J, Hogan AE, Toxavidis V, Balk SP, O'Shea D, O'Farrelly C & Exley MA 2012 Adipose tissue invariant NKT cells protect against diet-induced obesity and metabolic disorder through regulatory cytokine production. *Immunity* **37** 574–587. (doi:10.1016/j.immuni.2012.06.016)
- Mantell BS, Stefanovic-Racic M, Yang X, Dedousis N, Sipula IJ & O'Doherty RM 2011 Mice lacking NKT cells but with a complete complement of CD8⁺T cells are not protected against the metabolic abnormalities of diet-induced obesity. *PLoS ONE* 6 e19831. (doi:10.1371/journal.pone.0019831)

Mantovani A, Cassatella MA, Costantini C & Jaillon S 2011 Neutrophils in the activation and regulation of innate and adaptive immunity. *Nature Reviews. Immunology* **11** 519–531. (doi:10.1038/nri3024)

- Maury E & Brichard SM 2010 Adipokine dysregulation, adipose tissue inflammation and metabolic syndrome. *Molecular and Cellular Endocrinology* **314** 1–16. (doi:10.1016/j.mce.2009.07.031)
- Medzhitov R 2008 Origin and physiological roles of inflammation. *Nature* **454** 428–435. (doi:10.1038/nature07201)
- Meijer K, de Vos P & Priebe MG 2010 Butyrate and other short-chain fatty acids as modulators of immunity: what relevance for health? *Current Opinion in Clinical Nutrition and Metabolic Care* **13** 715–721. (doi:10.1097/MCO.0b013e32833eebe5)

- Molofsky AB, Nussbaum JC, Liang HE, Van Dyken SJ, Cheng LE, Mohapatra A, Chawla A & Locksley RM 2013 Innate lymphoid type 2 cells sustain visceral adipose tissue eosinophils and alternatively activated macrophages. *Journal of Experimental Medicine* **210** 535–549. (doi:10.1084/jem.20121964)
- Mortensen RF 2001 C-reactive protein, inflammation, and innate immunity. *Immunologic Research* **24** 163–176. (doi:10.1385/ IR:24:2:163)
- Mosser DM 2003 The many faces of macrophage activation. *Journal of Leukocyte Biology* **73** 209–212. (doi:10.1189/jlb.0602325)
- Mraz M, Lacinova Z, Drapalova J, Haluzikova D, Horinek A, Matoulek M, Trachta P, Kavalkova P, Svacina S & Haluzik M 2011 The effect of verylow-calorie diet on mRNA expression of inflammation-related genes in subcutaneous adipose tissue and peripheral monocytes of obese patients with type 2 diabetes mellitus. *Journal of Clinical Endocrinology* and Metabolism **96** E606–E613. (doi:10.1210/jc.2010-1858)
- Neels JG & Olefsky JM 2006 Inflamed fat: what starts the fire? *Journal of Clinical Investigation* **116** 33–35. (doi:10.1172/JCI27280)
- Nguyen MT, Favelyukis S, Nguyen AK, Reichart D, Scott PA, Jenn A, Liu-Bryan R, Glass CK, Neels JG & Olefsky JM 2007 A subpopulation of macrophages infiltrates hypertrophic adipose tissue and is activated by free fatty acids via Toll-like receptors 2 and 4 and JNK-dependent pathways. *Journal of Biological Chemistry* **282** 35279–35292. (doi:10.1074/jbc.M706762200)
- Nguyen KD, Qiu Y, Cui X, Goh YP, Mwangi J, David T, Mukundan L, Brombacher F, Locksley RM & Chawla A 2011 Alternatively activated macrophages produce catecholamines to sustain adaptive thermogenesis. *Nature* **480** 104–108. (doi:10.1038/nature10653)
- Nijhuis J, Rensen SS, Slaats Y, van Dielen FM, Buurman WA & Greve JW 2009 Neutrophil activation in morbid obesity, chronic activation of acute inflammation. *Obesity* **17** 2014–2018. (doi:10.1038/oby. 2009.113)
- Nishimura S, Manabe I, Nagasaki M, Eto K, Yamashita H, Ohsugi M, Otsu M, Hara K, Ueki K, Sugiura S *et al.* 2009 CD8⁺ effector T cells contribute to macrophage recruitment and adipose tissue inflammation in obesity. *Nature Medicine* **15** 914–920. (doi:10.1038/nm.1964)
- Odegaard JI, Ricardo-Gonzalez RR, Goforth MH, Morel CR, Subramanian V, Mukundan L, Red Eagle A, Vats D, Brombacher F, Ferrante AW *et al.* 2007 Macrophage-specific PPARγ controls alternative activation and improves insulin resistance. *Nature* **447** 1116–1120. (doi:10.1038/ nature05894)
- Oestreich KJ & Weinmann AS 2012 Master regulators or lineage-specifying? Changing views on CD4⁺T cell transcription factors *Nature Reviews*. *Immunology* **12** 799–804. (doi:10.1038/nri3321)
- Oh DY, Talukdar S, Bae EJ, Imamura T, Morinaga H, Fan W, Li P, Lu WJ, Watkins SM & Olefsky JM 2010 GPR120 is an omega-3 fatty acid receptor mediating potent anti-inflammatory and insulin-sensitizing effects. *Cell* **142** 687–698. (doi:10.1016/j.cell.2010.07.041)
- Ohashi K, Parker JL, Ouchi N, Higuchi A, Vita JA, Gokce N, Pedersen AA, Kalthoff C, Tullin S, Sams A *et al.* 2010 Adiponectin promotes macrophage polarization toward an anti-inflammatory phenotype. *Journal of Biological Chemistry* 285 6153–6160. (doi:10.1074/jbc.M109. 088708)
- Ohmura K, Ishimori N, Ohmura Y, Tokuhara S, Nozawa A, Horii S, Andoh Y, Fujii S, Iwabuchi K, Onoe K *et al.* 2010 Natural killer T cells are involved in adipose tissues inflammation and glucose intolerance in diet-induced obese mice. *Arteriosclerosis, Thrombosis, and Vascular Biology* **30** 193–199. (doi:10.1161/ATVBAHA.109.198614)
- Oliver E, McGillicuddy F, Phillips C, Toomey S & Roche HM 2010 The role of inflammation and macrophage accumulation in the development of obesity-induced type 2 diabetes mellitus and the possible therapeutic effects of long-chain n-3 PUFA. *Proceedings of the Nutrition Society* **69** 232–243. (doi:10.1017/S0029665110000042)
- O'Rahilly S 1997 Science, medicine, and the future. Non-insulin dependent diabetes mellitus: the gathering storm. *BMJ* **314** 955–959. (doi:10.1136/ bmj.314.7085.955)

- Osborn O & Olefsky JM 2012 The cellular and signaling networks linking the immune system and metabolism in disease. *Nature Medicine* **18** 363–374. (doi:10.1038/nm.2627)
- Pacifico L, Di Renzo L, Anania C, Osborn JF, Ippoliti F, Schiavo E & Chiesa C 2006 Increased T-helper interferon-γ-secreting cells in obese children. *European Journal of Endocrinology* **154** 691–697. (doi:10.1530/eje.1. 02138)
- Patsouris D, Li PP, Thapar D, Chapman J, Olefsky JM & Neels JG 2008 Ablation of CD11c-positive cells normalizes insulin sensitivity in obese insulin resistant animals. *Cell Metabolism* 8 301–309. (doi:10.1016/ j.cmet.2008.08.015)
- Prieur X, Mok CY, Velagapudi VR, Nunez V, Fuentes L, Montaner D, Ishikawa K, Camacho A, Barbarroja N, O'Rahilly S *et al.* 2011 Differential lipid partitioning between adipocytes and tissue macrophages modulates macrophage lipotoxicity and M2/M1 polarization in obese mice. *Diabetes* **60** 797–809. (doi:10.2337/db10-0705)
- Rausch ME, Weisberg S, Vardhana P & Tortoriello DV 2008 Obesity in C57BL/6J mice is characterized by adipose tissue hypoxia and cytotoxic T cell infiltration. *International Journal of Obesity* **32** 451–463. (doi:10.1038/sj.ijo.0803744)
- Ravussin E & Smith SR 2002 Increased fat intake, impaired fat oxidation, and failure of fat cell proliferation result in ectopic fat storage, insulin resistance, and type 2 diabetes mellitus. *Annals of the New York Academy* of Sciences **967** 363–378. (doi:10.1111/j.1749-6632.2002.tb04292.x)
- Ricardo-Gonzalez RR, Red Eagle A, Odegaard JI, Jouihan H, Morel CR, Heredia JE, Mukundan L, Wu D, Locksley RM & Chawla A 2010 IL-4/STAT6 immune axis regulates peripheral nutrient metabolism and insulin sensitivity. *PNAS* **107** 22617–22622. (doi:10.1073/pnas. 1009152108)
- Rocha VZ, Folco EJ, Sukhova G, Shimizu K, Gotsman I, Vernon AH & Libby P 2008 Interferon-γ, a Th1 cytokine, regulates fat inflammation: a role for adaptive immunity in obesity. *Circulation Research* **103** 467–476. (doi:10.1161/CIRCRESAHA.108.177105)
- Rolff J & Siva-Jothy MT 2003 Invertebrate ecological immunology. *Science* **301** 472–475. (doi:10.1126/science.1080623)
- Rosenberg HF, Dyer KD & Foster PS 2013 Eosinophils: changing perspectives in health and disease. *Nature Reviews. Immunology* **13** 9–22. (doi:10.1038/nri3341)
- Roubicek T, Bartlova M, Krajickova J, Haluzikova D, Mraz M, Lacinova Z, Kudla M, Teplan V & Haluzik M 2009 Increased production of proinflammatory cytokines in adipose tissue of patients with end-stage renal disease. *Nutrition* 25 762–768. (doi:10.1016/j.nut.2008.12.012)
- Rusten TE, Lindmo K, Juhasz G, Sass M, Seglen PO, Brech A & Stenmark H 2004 Programmed autophagy in the *Drosophila* fat body is induced by ecdysone through regulation of the PI3K pathway. *Developmental Cell* **7** 179–192. (doi:10.1016/j.devcel.2004.07.005)
- Saberi M, Woods NB, de Luca C, Schenk S, Lu JC, Bandyopadhyay G, Verma IM & Olefsky JM 2009 Hematopoietic cell-specific deletion of toll-like receptor 4 ameliorates hepatic and adipose tissue insulin resistance in high-fat-fed mice. *Cell Metabolism* **10** 419–429. (doi:10.1016/j.cmet.2009.09.006)
- Satoh M, Andoh Y, Clingan CS, Ogura H, Fujii S, Eshima K, Nakayama T, Taniguchi M, Hirata N, Ishimori N *et al.* 2012 Type II NKT cells stimulate diet-induced obesity by mediating adipose tissue inflammation, steatohepatitis and insulin resistance. *PLoS ONE* **7** e30568. (doi:10.1371/journal.pone.0030568)
- Schipper HS, Prakken B, Kalkhoven E & Boes M 2012a Adipose tissueresident immune cells: key players in immunometabolism. *Trends* in Endocrinology and Metabolism 23 407–415. (doi:10.1016/j.tem. 2012.05.011)
- Schipper HS, Rakhshandehroo M, van de Graaf SF, Venken K, Koppen A, Stienstra R, Prop S, Meerding J, Hamers N, Besra G *et al.* 2012b Natural killer T cells in adipose tissue prevent insulin resistance. *Journal of Clinical Investigation* **122** 3343–3354. (doi:10.1172/JCI62739)
- Shaul ME, Bennett G, Strissel KJ, Greenberg AS & Obin MS 2010 Dynamic, M2-like remodeling phenotypes of CD11c+ adipose tissue

http://joe.endocrinology-journals.org DOI: 10.1530/JOE-14-0283 © 2014 Society for Endocrinology Printed in Great Britain Published by Bioscientifica Ltd.

macrophages during high-fat diet-induced obesity in mice. *Diabetes* **59** 1171–1181. (doi:10.2337/db09-1402)

- Shoelson SE, Lee J & Yuan M 2003 Inflammation and the IKKβ/IκB/NF-κB axis in obesity- and diet-induced insulin resistance. *International Journal of Obesity and Related Metabolic Disorders* **27**(Suppl 3) S49–S52. (doi:10.1038/sj.ijo.0802501)
- Solinas G, Vilcu C, Neels JG, Bandyopadhyay GK, Luo JL, Naugler W, Grivennikov S, Wynshaw-Boris A, Scadeng M, Olefsky JM *et al.* 2007 JNK1 in hematopoietically derived cells contributes to diet-induced inflammation and insulin resistance without affecting obesity. *Cell Metabolism* **6** 386–397. (doi:10.1016/j.cmet.2007.09.011)
- Sondergaard L 1993 Homology between the mammalian liver and the *Drosophila* fat body. *Trends in Genetics* **9** 193. (doi:10.1016/0168-9525(93)90113-V)
- Spencer LA & Weller PF 2010 Eosinophils and Th2 immunity: contemporary insights. *Immunology and Cell Biology* **88** 250–256. (doi:10.1038/icb.2009.115)
- Stefanovic-Racic M, Yang X, Turner MS, Mantell BS, Stolz DB, Sumpter TL, Sipula IJ, Dedousis N, Scott DK, Morel PA *et al.* 2012 Dendritic cells promote macrophage infiltration and comprise a substantial proportion of obesity-associated increases in CD11c+ cells in adipose tissue and liver. *Diabetes* **61** 2330–2339. (doi:10.2337/db11-1523)
- Steinman RM 2008 Dendritic cells *in vivo*: a key target for a new vaccine science. *Immunity* **29** 319–324. (doi:10.1016/j.immuni.2008.08.001)
- Stienstra R, Duval C, Keshtkar S, van der Laak J, Kersten S & Muller M 2008 Peroxisome proliferator-activated receptor γ activation promotes infiltration of alternatively activated macrophages into adipose tissue. *Journal of Biological Chemistry* **283** 22620–22627. (doi:10.1074/jbc. M710314200)
- Sun K, Kusminski CM & Scherer PE 2011 Adipose tissue remodeling and obesity. *Journal of Clinical Investigation* **121** 2094–2101. (doi:10.1172/ JCI45887)
- Sun S, Ji Y, Kersten S & Qi L 2012 Mechanisms of inflammatory responses in obese adipose tissue. *Annual Review of Nutrition* **32** 261–286. (doi:10.1146/annurev-nutr-071811-150623)
- Talukdar S, Oh da Y, Bandyopadhyay G, Li D, Xu J, McNelis J, Lu M, Li P, Yan Q, Zhu Y *et al.* 2012 Neutrophils mediate insulin resistance in mice fed a high-fat diet through secreted elastase. *Nature Medicine* **18** 1407–1412. (doi:10.1038/nm.2885)
- Trachta P, Dostalova I, Haluzikova D, Kasalicky M, Kavalkova P, Drapalova J, Urbanova M, Lacinova Z, Mraz M & Haluzik M 2014 Laparoscopic sleeve gastrectomy ameliorates mRNA expression of inflammationrelated genes in subcutaneous adipose tissue but not in peripheral monocytes of obese patients. *Molecular and Cellular Endocrinology* **383** 96–102. (doi:10.1016/j.mce.2013.11.013)
- Trayhurn P 2013 Hypoxia and adipose tissue function and dysfunction in obesity. *Physiological Reviews* **93** 1–21. (doi:10.1152/physrev.00017. 2012)
- Uysal KT, Wiesbrock SM, Marino MW & Hotamisligil GS 1997 Protection from obesity-induced insulin resistance in mice lacking TNF- α function. *Nature* **389** 610–614. (doi:10.1038/39335)
- Vandanmagsar B, Youm YH, Ravussin A, Galgani JE, Stadler K, Mynatt RL, Ravussin E, Stephens JM & Dixit VD 2011 The NLRP3 inflammasome instigates obesity-induced inflammation and insulin resistance. *Nature Medicine* 17 179–188. (doi:10.1038/nm.2279)
- Vasudevan AR, Wu H, Xydakis AM, Jones PH, Smith EO, Sweeney JF, Corry DB & Ballantyne CM 2006 Eotaxin and obesity. *Journal of Clinical Endocrinology and Metabolism* **91** 256–261. (doi:10.1210/jc.2005-1280)
- Vitseva OI, Tanriverdi K, Tchkonia TT, Kirkland JL, McDonnell ME, Apovian CM, Freedman J & Gokce N 2008 Inducible Toll-like receptor and NF-kB regulatory pathway expression in human adipose tissue. *Obesity* 16 932–937. (doi:10.1038/oby.2008.25)

- Weisberg SP, McCann D, Desai M, Rosenbaum M, Leibel RL & Ferrante AW Jr 2003 Obesity is associated with macrophage accumulation in adipose tissue. *Journal of Clinical Investigation* **112** 1796–1808. (doi:10.1172/ICI200319246)
- Weisberg SP, Hunter D, Huber R, Lemieux J, Slaymaker S, Vaddi K, Charo I, Leibel RL & Ferrante AW Jr 2006 CCR2 modulates inflammatory and metabolic effects of high-fat feeding. *Journal of Clinical Investigation* **116** 115–124. (doi:10.1172/JCI24335)
- Williamson RT 1901 On the treatment of glycosuria and diabetes mellitus with sodium salicylate. *BMJ* **1** 760–762. (doi:10.1136/bmj.1.2100.760)
- Winer S, Chan Y, Paltser G, Truong D, Tsui H, Bahrami J, Dorfman R, Wang Y, Zielenski J, Mastronardi F *et al.* 2009 Normalization of obesityassociated insulin resistance through immunotherapy. *Nature Medicine* **15** 921–929. (doi:10.1038/nm.2001)
- Winer DA, Winer S, Shen L, Wadia PP, Yantha J, Paltser G, Tsui H, Wu P, Davidson MG, Alonso MN *et al.* 2011 B cells promote insulin resistance through modulation of T cells and production of pathogenic IgG antibodies. *Nature Medicine* **17** 610–617. (doi:10.1038/nm.2353)
- Wong N, Fam BC, Cempako GR, Steinberg GR, Walder K, Kay TW, Proietto J & Andrikopoulos S 2011 Deficiency in interferon-γ results in reduced body weight and better glucose tolerance in mice. *Endocrinology* **152** 3690–3699. (doi:10.1210/en.2011-0288)
- Wu D, Molofsky AB, Liang HE, Ricardo-Gonzalez RR, Jouihan HA, Bando JK, Chawla A & Locksley RM 2011 Eosinophils sustain adipose alternatively activated macrophages associated with glucose homeostasis. *Science* **332** 243–247. (doi:10.1126/science.1201475)
- Wu L, Parekh VV, Gabriel CL, Bracy DP, Marks-Shulman PA, Tamboli RA, Kim S, Mendez-Fernandez YV, Besra GS, Lomenick JP *et al.* 2012 Activation of invariant natural killer T cells by lipid excess promotes tissue inflammation, insulin resistance, and hepatic steatosis in obese mice. *PNAS* **109** E1143–E1152. (doi:10.1073/pnas.1200498109)
- Xu H, Barnes GT, Yang Q, Tan G, Yang D, Chou CJ, Sole J, Nichols A, Ross JS, Tartaglia LA *et al.* 2003 Chronic inflammation in fat plays a crucial role in the development of obesity-related insulin resistance. *Journal of Clinical Investigation* **112** 1821–1830. (doi:10.1172/JCI200319451)
- York DA, Rossner S, Caterson I, Chen CM, James WP, Kumanyika S, Martorell R, Vorster HH & American Heart A 2004 Prevention Conference VII: obesity, a worldwide epidemic related to heart disease and stroke: Group I: worldwide demographics of obesity. *Circulation* **110** e463–e470. (doi:10.1161/01.CIR.0000140125.26161.49)
- Yuan M, Konstantopoulos N, Lee J, Hansen L, Li ZW, Karin M & Shoelson SE 2001 Reversal of obesity- and diet-induced insulin resistance with salicylates or targeted disruption of Ikkβ. *Science* 293 1673–1677. (doi:10.1126/science.1061620)
- Zeyda M, Farmer D, Todoric J, Aszmann O, Speiser M, Gyori G, Zlabinger GJ & Stulnig TM 2007 Human adipose tissue macrophages are of an antiinflammatory phenotype but capable of excessive pro-inflammatory mediator production. *International Journal of Obesity* **31** 1420–1428. (doi:10.1038/sj.ijo.0803632)
- Zhang N & Bevan MJ 2011 CD8(+) T cells: foot soldiers of the immune system. *Immunity* **35** 161–168. (doi:10.1016/j.immuni.2011.07.010)
- Zhu L, Wu Y, Wei H, Xing X, Zhan N, Xiong H & Peng B 2011 IL-17R activation of human periodontal ligament fibroblasts induces IL-23 p19 production: differential involvement of NF-κB versus JNK/AP-1 pathways. *Molecular Immunology* **48** 647–656. (doi:10.1016/j.molimm.2010. 11.008)
- Zuniga LA, Shen WJ, Joyce-Shaikh B, Pyatnova EA, Richards AG, Thom C, Andrade SM, Cua DJ, Kraemer FB & Butcher EC 2010 IL-17 regulates adipogenesis, glucose homeostasis, and obesity. *Journal of Immunology* 185 6947–6959. (doi:10.4049/jimmunol.1001269)

Received in final form 3 July 2014 Accepted 8 July 2014 Accepted Preprint published online 8 July 2014

Journal of Endocrinology